MECHANISM OF CONTAMINATION WITH AFLATOXINS OF COTTON SEEDS AND PRODUCTS OF THEIR PROCESSING

> G. V. Tarasyuk, M. T. Tutakhozaev, R. A. Mirkina, UI and T. T. Shakirov

A possible mechanism of the contamination of cotton seeds and the products of their processing with aflatoxins has been studied. It has been found that the contamination of cotton seeds with aflatoxin takes place mainly on storage in the cotton ginning factory and the oils and fats combine. In the processing of the cotton seeds, part of the aflatoxins passes into the final products and part decomposes. Meal stored in the oil and fats combine undergo secondary contamination with aflatoxins.

It is known that the development of <u>Aspergillus flavus</u> fungi – the producing agents of aflatoxins, which possess hepatoxic and hepatocarcinogenic properties [1, 2] – takes place under certain conditions in various food products [3] and feedstuffs [4] and also in oil crops, including cotton seeds and the products of their processing [5].

Experi-	Soil sa	umpie	Amou	Amount of aflatoxin, µg/kg					
ment No.	month takén (1985)	depth, cm	B,	C,	B,	C,			
1 2 3 <b>4</b> 5	iV	015	0.32 0.07 0.25 0.15 0.20	0.32 0,07 0.25 0.15 0,20	0.15 0,04 0.13 0,10 0,09	0,15 0,04 0,13 0,10 0,09			
6 7 8 9 10		15-30	0,20 0,10 0,15 0,05 0,02	0,20 0,10 0,15 0,05 0,62	0.10 0.05 0.07 0.04 0.04	0,10 0,05 0,07 0,04 0,04			
11 12 13 14 15	VI	0-15	0.12 0.20 0.10 0.09 0.10	0,12 0,20 0,10 0,09 0,10	0,06 0.10 0,19 0,09 0,08	0,06 0,10 0,10 0,09 0,08			
16 17 18 19 20		15-30	0,07	0,07 	0,64	0,04			
21 22 23 24 25	IX	015	0,20 0,40 0,25 0,25 0,25 0,32	0,20 0,40 0,25 0,25 0,25 0,32	0,10 0,20 0,18 0,12 0,10	0,10 0,20 0,18 0,12 0,10			
26 27 28 29 30		1530	0,04 	0,04 					
31 35	XI			Not dete	cted	•			

TABLE 1. Amounts of Aflatoxins in Samples of Soil

Institute of the Chemistry of Plant Substances, Academy of Sciences of the Uzbek SSR, Tashkent. Translated from Khimiya Prirodnykh Soedinenii, No. 1, pp. 118-123, January-February, 1988. Original article submitted June 2, 1987.

TABLE 2. Amounts of Aflatoxins in the Cotton Plant

Sample	Time of taking	Amo	unt of afla	toxin, μg/k	g
No.		B <sub>1</sub>	C,	B <sub>2</sub>	C,
1 2 3 4 5 6 7 8 9 10	15-Day shoots Budding and flower- ing phase	0,70 0,60 0,51 0,46 0,62 0,50 	0,70 0,60 0,51 0,46 0,62 0,50 	0,35 0,30 0,25 0,23 0,30 0,25	0,35 0,30 0,25 0,23 0,30 0,25  
11 50	Fruit-forming phase	Not	detected	l	1

TABLE 3. Amounts of Aflatoxins in Seeds Stored for a Month at a MZhK

Comp 1 of	Depth of	Amou	nt of aflato	xins, µg/k	g
Sample No.	sampling,	В,	C <sub>1</sub>	Bs	C,
1 2 3 4	20	0,42 0,22 0,15 0,30	0,42 0,22 0,15 0,30	0,21 0,11 0,08 0,15	0,21 0,11 0,08 0,15
5 6 7 8	50	0,35 0,10 0,18 0,20	0,35 0,10 0,18 0,20	0,18 0,05 0,09 0,10	0,18 0,05 0,09 0,10
9 10 11 12	1000	0,21 0,09 0,09 0,10	0,21 0,09 0,09 0,10	0,11 0,05 0,05 0,05	$\begin{array}{c} 0,11\\ 0,05\\ 0,05\\ 0,05\\ 0,05\end{array}$

TABLE 4. Amounts of Aflatoxins in Seeds Stored for 3 Months in a MZhK

		0	pen me	thod		Clo	sed me	thod	
Sample No.	Depth of sampling,			Αποι	int of	aflat	oxins,	μg/k	g
	cm	B,	Ci	Ba	С,	B <sub>1</sub>	C,	81	C,
1 2 3 4 5 6	20	0,85 1,23 0,57 3,00 0,78	0,85 0,15 0,09 0,57 3,00 0,38	$\begin{array}{r} 0,42 \\ \\ 0,61 \\ 0,50 \\ 1,50 \\ 0,19 \end{array}$	0,42 0,07 0,05 0,28 1,50 0,19	2,50 1,90 1,13 3,00 2,55 0,81	1,2) 1,9) 1,85 1,50 1,28	1,25 0,95 - 0,56 0,72 -	0,6) 0,95 
7 8 9 10 11	50	0,19 0,73 0,21 1,10 1,35	0,55 0,21 $\overline{0,77}$	$ \begin{array}{c}         0,35 \\         0,22 \\         - \\         0,48     \end{array} $	0,27 0,11 0,38	0,50 0,19 2,40 1,90 0,70	0,50 0,19 2,40 1,35 0,90	0,25 0,38 1,20 1,47 0,50	0,25 0,09 1,20 1,49 0,50
12 13 14 15 16	1000	0,15 0,15 0,30 0,15 0,14	0,15 0,15 0,30 0,15 0,14	0.07 0.07 0.15 0.07 0.07	0.07 0.07 0.30 0.07 0.07				

We have previously [6] reported the results of the determination of the amount of aflatoxins in cottonseed meal and some samples of protein obtained from it. In the present paper we give the results of a study of a possible mechanism of the contamination of cottonseed meal with the products of the vital activity of <u>Aspergillus flavus</u> fungi.

Cotton seeds before sowing, samples of soil, 15- and 50-day cotton-plant shoots (epigeal part), the seeds of a ripe crop, and samples of crushed seeds, meal, and refined oil were investigated. Samples were taken from control sections of the experimental fields of the Institute of the Chemistry of Plant Substances in the Kommunizm kholkhoz [State farm] and in the Tashkent oils and fats combine (MZhK) in 1985.

					·			Amount of	f aflatox	Amount of aflatoxins, µg/kg	kg					-			
n th	in the crushed seeds	ed see	spa	ir	in the seed husks	d husks			n the mea he extrac	in the meal from the extraction shop		in loa	the mea ding st	in the meal from the loading station	che	in	in the refined oil	ned oil	
В,	J	B,	ۍ	В,	ŭ	B,	ۍ	B,	Ű	l B,	ۍ ا	B,	 ט	B,	ن	B	Ū	B,	ڻ
3-1-58	2,08 0,69 0,95 3,15	1,04 0,35 0,50 2,07	1,04 0,50 2,07	0,000 0,000 80 80 80 80 80 80 80 80 80 80 80 80	0,40 0,30 0,45 0,45	0,31 0,35 0,72 0,50	0,50 5,33 0,50 0,50 0,50 0,50 0,50	2,00 2,60 2,60 2,00 2,00 2,00	0,93 0,24 0,78 0,78 2,17	0,85 0,10 0,50 1,54 1,54	0,85 0,10 0,40 1,50	1,74 0,50 0,50 0,90	5.02 0.50 5.02 0.50 0.50	3,00 3,00 3,00	3-1-0-0-0-0-0-0-0-0-0-0-0-0-0-0-0-0-0-0-	00000 40000 44000	00000 442000	00000 00000 00000	00000 00000 00000

See
Cotton
5
<b>Products</b>
Processing ]
the
in
Aflatoxins
of
Amounts o
<u>с</u> .
ABLE
AB

In none of the samples investigated did the level of aflatoxins exceed the maximum acceptable concentration (MAC) laid down, which agrees with results obtained by Kyuz [7, 8]. The MAC for aflatoxin  $B_1$  (the most toxic variety [9]) corresponds to 5  $\mu$ g/kg for all food products [10].

No aflatoxins were detected in seeds taken for analysis before sowing, which can apparently be explained by their treatment with fungicidal preparations.

It is also interesting to note that samples of soil taken in November contained no aflatoxins, although they were detected in the preceding months (Table 1). This is possibly connected with the irrigation period [11].

Table 2 gives the results of a study of 50 samples of the epigeal part of the cotton plant.

With the growth of the plant, the concentration of aflatoxins in it fell. While in the epigeal part of 15-day shoots the concentrations of aflatoxins  $B_1$ ,  $B_2$ ,  $C_1$ , and  $C_2$  were comparatively high, no aflatoxins were detected in the mature plant (leaves, stems, unopened bolls, fibers, seeds). The variation in the amounts of aflatoxin at the end of the vegetation of the cotton plant is connected with the nature of the vital activity of the <u>Aspergillus</u> fungi [11], i.e., conditions not favorable for the development of these fungi inhibit their vital activity.

Thus, aflatoxins pass from the soil into the plant and then with the growth of the plant their amount falls and ripe cotton seeds contain none of them.

Cotton seeds after harvesting are stored for an average of 3 months at the collecting station and the cotton-ginning factory, and are then transferred to MZhK. We analyzed seeds stored in the MZhK by the open method — in bales — and found that aflatoxins had accumulated in small amounts mainly in the upper layers (Table 3).

With an increase in the time of storage to 3 months there was a vigorous development of the <u>Aspergillus flavus</u> fungi and, correspondingly, of their metabolites, aflatoxins (Table 4), the level of contamination of the seeds stored by the open and closed methods being practically the same, which, in all probability, is explained by the use of active ventilation on storage.

We simultaneously investigated the products of the processing of cotton seeds (Table 5).

The amount of aflatoxins in the crushed seeds ranged from 0.69 to 2.08  $\mu$ g/kg for B<sub>1</sub> and C<sub>1</sub> and from 0.35 to 1.04  $\mu$ g/kg for B<sub>2</sub> and C<sub>2</sub>, while in the meal from the extraction shops the amount of aflatoxins had fallen under the action of the heat treatment and the gasoline extraction of the initial product. However, in samples kept in a store and intended for the working up of the meal, the amount of aflatoxins had increased, i.e., there was secondary contamination with aflatoxins. Some of the aflatoxins passed in the micellae, and in the final product – the refined oil – they were found at levels of 0.04  $\mu$ g/kg for B<sub>1</sub> and C<sub>1</sub> and 0.02  $\mu$ g/kg for B<sub>2</sub> and C<sub>2</sub>, which are considerably below the MAC.

Thus, freshly gathered seeds contain no aflatoxins but subsequently, on processing in the cotton-ginning factories, they undergo invasion by the <u>Aspergillus</u> fungi the development of which depends on the conditions and nature of storage. The aflatoxins detected in the cotton seeds and meal after preliminary storage are the result of secondary contamination.

## EXPERIMENTAL

<u>The amounts of aflatoxins B<sub>1</sub>, C<sub>1</sub>, B<sub>2</sub>, and C<sub>2</sub> in the soil were determined by the method of [10] with some modifications. A sample (100 g) of soil that had been ground and passed through a sieve with apertures having a diameter of 1 mm was extracted with an 80% solution of acetone by shaking for 30 min, and the mixture was filtered. To 50 ml of the filtrate was added 25 ml of a 15% solution of lead acetate, and the resulting precipitate was filtered off. Of this filtrate, 80 ml was transferred to a separatory funnel, and 40 ml of hexane was added, after which the aqueous acetone layer was separated off. The aflatoxins were extracted from the aqueous acetone solution successively with 40 ml of chloroform and with chloroform—acetone (3:1). The combined extracts were dried with sodium sulfate and evaporated in a rotary evaporator in vacuum to dryness at a temperature not exceeding 60°C. The residue was dissolved in 0.1 ml of chloroform and the solution was analyzed with the aid of two-dimensional TLC: first direction, ether; second direction, chloroform—acetone (9:1).</u>

The detection in UV light of spots corresponding in their chromatographic mobilities and the color of their fluorescence to the spots of aflatoxin standards showed the presence of aflatoxins in the samples investigated. Confirmatory tests were carried out by spraying the TLC plates with a solution of nitric acid in water (1:2). The aflatoxins were determined quantitatively by matching corresponding dilutions, which made it possible to trap on the plate spots with vanishingly low fluorescence. According to the literature [12], such spots contain 0.004  $\mu$ g/kg of B<sub>1</sub> and 0.0002  $\mu$ g/kg of C<sub>2</sub>, which enables the amount of aflatoxin in the sample to be calculated.

<u>The amounts of aflatoxins  $B_1$ ,  $C_1$ ,  $B_2$ , and  $C_2$  in cotton-plant shoots were determined as described above with additional purification, consisting in three treatments of the aqueous acetone extract with hexane to eliminate pigments.</u>

<u>The amounts of aflatoxins  $B_1$ ,  $C_1$ ,  $B_2$ , and  $C_2$  in cotton seeds and in the meal obtained from them were determined after additional purification of the aqueous acetone extract from lipids on a chromatographic column containing silica gel.</u>

<u>Determination of the Amounts of Aflatoxins  $B_1$ ,  $C_1$ ,  $B_2$ , and  $C_2$  in the Cottonseed Oil. The aflatoxins were extracted by a published method [13]. The extract of aflatoxins was analyzed by the method described above with additional chromatographic purification using hexane.</u>

## EXPERIMENTAL

The investigations have shown that the contamination of the cotton seeds with aflatoxin takes place mainly on storage in the cotton-ginning factory and the oils and fats combine (MZhK).

In the processing of cotton seeds, part of the aflatoxins passes into the final products and part decomposes. Meal stored in the MZhK undergoes secondary contamination with aflatoxins.

## LITERATURE CITED

- 1. W. H. Butler and J. M. Barnes, Mycotoxins, Elsevier, New York (1974), p. 2.
- G. N. Wogan, in: Methods in Cancer Research, M. Buch (ed.), Academic Press, New York (1973), p. 309.
- 3. T. C. Campbell and Z. Stoloff, J. Agric. Food Chem., <u>22</u>, No. 6, 1006 (1974).
- 4. L. Stoloff, in: Mycotoxins and Other Fungal Related Food Problems, J. V. Rodricks (ed.), American Chemical Society, Washington, DC (1976), p. 23.
- P. B. Marsh, M. E. Simpson, G. O. Craig, J. Donoso, and H. H. Ramey, J. Environ. Qual., <u>2</u>, 276 (1973).
- 6. G. V. Tarasyuk, M. T. Turakhozhaev, T. T. Shakirov, and R. A. Mirkina, Khim. Prir. Soedin., 843 (1985).
- 7. E. P. Kyuz, S. N. Burnasheva, and V. Ya. Stoikova, Maslov.-Zhir. Prom-st', 13 (1974).
- 8. K. G. Koshlakova, E. P. Kyuz, and V. Ya. Stoikova, Maslov.-Zhir. Prom-st', 11 (1983). 9. A. A. Pokrovskii, L. V. Krachchenko, and V. A. Tutel'yan, Toxicology. Advances in
- Sciences and Technology Series [in Russian], VINITI, Moscow, Vol. 8 (1977), p. 15.
- Methodological Recommendation on the Detection, Identification, and Determination of Aflatoxins in Food Products [in Russian], Moscow (1981).
- 11. WHO Task Group on Environmental Health Criteria, Mycotoxins. WHO, Geneva (1979).
- 12. Z. S. Lvova and O. D. Doronina, Microdetection of Mycotoxins in Grain Extracts and Fungi Cultures, Center of International Projects, GKNT, Moscow (1984).
- B. Letutour, I. Tantaoui-Elaraki, and L. Ihlal, J. Am. Oil Chem. Soc., <u>60</u>, No. 4, 835 (1983).